Structure Determination of LL-F28249 α , β , γ , and λ , Potent Antiparasitic Macrolides from *Streptomyces cyaneogriseus* ssp. *noncyanogenus*

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The structures of four antiparasitic macrocyclic lactones, LL-F28249 α , λ , β , and γ (1)—(4) derived from the actinomycete *Streptomyces cyaneogriseus* ssp. *noncyanogenus*, were determined by spectroscopic methods and by *X*-ray crystallography of the γ -component (4).

Structures have been determined for a new family of potent antiparasitic antibiotics derived from *Streptomyces cyaneo*griseus ssp. noncyanogenus. These compounds designated LL-F28249 α , λ , β , and γ (1)—(4) possess potent activity against commercially important endo- and ecto-parasites of cattle, sheep, swine, dogs, and horses.[†] The LL-F28249 compounds are related to the milbemycins¹ and avermectins² but are unique in that they each contain a trisubstituted double



[†] The spectrum of antiparasitic activity demonstrated by these compounds is similar to that obtained with the avermectin derivative ivermectin. Details concerning antiparasitic efficacy will be published separately.

bond (Δ^{26}) in their side chains. This feature, combined with the rare substitution pattern for C-22 and -23 (found only in the avermectin '2' series)³ may be responsible for their highly potent parasiticidal activity.

LL-F28249 α (1) was determined by high-resolution electron-impact mass spectrometry (h.r.e.i.m.s.) to have the formula C₃₆H₅₂O₈.‡ Its characteristic u.v. chromophore and e.i.m.s. fragment ions at m/z 151, 248, and 314 were clear indicators of the basic milbemycin structure.¹ Comparison of ¹H and ¹³C n.m.r. data with those of the known milbemycins indicated identity between (1) and milbemycin D (5) for C-1—21, comprising the macrocycle and left-hand portion of

 $[\]pm F28249\alpha$ (1): C₃₆H₅₂O₈ (*M*+ *m*/*z* 612.3705, calc. 612.3662); $[\alpha]_{D}^{26}$ +133° (c 0.3, acetone); λ_{max} . (MeOH) (log ε) 244 (4.47) nm; v_{max} . (KBr) 3439, 2960, 2925, 1714, 1454, 1374, 1338, 1171, 1120, 996, and 967 cm⁻¹; m/z (electron impact, 70 eV) 612 (M^+ ; 16%), 594 (17), 466 (55), 448 (17), 425 (15), 354 (30), 314 (27), 248 (13), 247 (12), 237 (11), 219 (13), 204 (17), 152 (88), 151 (93), and 95 (100); ¹H n.m.r. (CDCl₃, 300 MHz), $\delta(Me_4Si)$ (numbering corresponds to that in Figure 2) 0.802 (3H, d, J 5.9 Hz, Me-24a), 0.883 (1H, m, 18-Hax), 0.953 (3H, d, J 6.6 Hz, Me-29 or -28a), 1.00 (3H, d, J 6.6 Hz, Me-28a or -29), 1.05 (3H, d, J 6.6 Hz, Me-12a), 1.40 (1H, t, J 12 Hz, 20-H_{ax}), 1.53 (3H, s, Me-14a), 1.61 (3H, s, Me-26a), 1.70 (1H, dd, J 14, 3.2 Hz, 22-H_{ax}), 1.87 (3H, br.s, Me-4a), 1.99 (1H, dd, J 14, 2.4 Hz, 22-Hea), 2.58 (1H, m, 28-H), 3.27 (1H, q, J 2.0 Hz, 2-H), 3.62 (1H, m, 17-H), 3.74 (1H, d, J 11 Hz, 25-H), 3.80 (1H, m, 23-H), 3.95 (1H, d, J 6.2 Hz, 6-H), 4.29 (1H, br.d, J 6.0 Hz, 5-H), 4.68 (2H, AB, CH₂-8a), 4.95 (1H, dd, J 11, 5 Hz, 15-H), 5.20 (1H, d, J 9.1 Hz, 27-H), and 5.41 (1H, br.s, 3-H); ¹³C n.m.r. (CDCl₃, 75 MHz), δ(Me₄Si), 11.0 (26a), 13.9 (24a), 15.5 (14a), 19.9 (4a), 22.2 (12a), 22.8 (28a, 29 unresolved), 26.8 (28), 34.7 (16), 35.9 (24), 36.0 (12), 36.1 (18), 40.7 (20), 41.1 (22), 45.7 (2), 48.4 (13), 67.7 (17), 67.8 (5), 68.4 (8a), 68.5 (19), 69.3 (23), 76.7 (25), 79.3 (6), 80.2 (7), 99.7 (21), 118.0 (3), 120.3 (9, 15 unresolved), 123.3 (10), 130.6 (26), 137.2 (27), 137.3 (14), 137.7 (4), 139.4 (8), 142.8 (11), and 173.4 (1).

(5) (6)





Figure 1. E.i.m.s. fragmentation of LL-F28249 α (1).

the spiroacetal system. The remaining portion of the molecule, including the right side of the spiro system, consists of the fragment $C_{11}H_{20}O_2$. Hydroxy substitution of this ring was suggested by observation of several ions in the e.i. mass spectrum of (1) which indicated an additional unsaturation not present in the parent molecule (Figure 1). One of these, m/z482.2648 ($C_{29}H_{38}O_6$), corresponds to loss of the side chain plus C-25 and the attached oxygen. This ion presumably arises through dehydration to the Δ^{22} compound followed by retro-Diels-Alder fragmentation of the ring, demanding



Figure 2. ORTEP drawing of LL-F28249y (4).

hydroxy substitution at either the 22 or the 23 position. Evidence for 23-hydroxy substitution comes from the formation of the m/z 442.2375 ($C_{26}H_{34}O_6$) ion resulting from loss of C-23—C-29 including 2 oxygens. The 23-hydroxy promotes the cleavage of the C-22–C-23 bond and the fragmentation is completed by fission of the C-21–O bond.

The structure of the novel six-carbon side chain of (1) was determined by analysis of ¹H and ¹³C n.m.r. data. The trisubstituted double bond was readily identified from ¹³C n.m.r. signals at δ 130.6 (s) and 137.2 (d). The position of the isopropyl and methyl substituents about the double bond was assured by the observed 9 Hz doublet for 27-H at δ 5.20, indicating vicinal coupling to the isopropyl methine.

LL-F28249 λ (2) was readily recognized as a methoxy derivative of (1) by characteristic ¹H (δ 3.50, 3H, s) and ¹³C (δ 51.7, q) n.m.r. signals for the *O*-methyl group. Location of the methyl on the 5-oxygen was established from the appearance of appropriately shifted e.i.m.s. fragment ions. Similar arguments were applied for the structure assignments of the β (3) and γ components (4), which contain a terminal methyl substituent on the side chain in place of the isopropyl group of (1) and (2). In the ¹H n.m.r. spectra of (3) and (4) the proton on the double bond in the side chain gives rise to a doublet of quartets showing vicinal coupling (6.5–7.0 Hz) to the terminal methyl and allylic coupling (1.0–1.2 Hz) to the 25-H.

Slow evaporation of methanol-water solutions obtained from reversed-phase chromatography yielded crystals of LL-F28249 γ (4), which were used for X-ray crystallography.§ The resulting structure is given in Figure 2. As expected from n.m.r. data, the relative stereochemistry of all substituents is identical to that found for avermectin B_{2b} (6).⁴ In addition, the Δ^{26} double bond was clearly shown to have the *trans*configuration.

Preliminary results from biosynthetic experiments indicated identical labelling patterns with acetate and propionate in the macrocycle and spiroacetal rings as was found for milbemycin D^5 and avermectin.⁶ The structure of the side chains in (1)—(4), however, reflects a fundamental difference in their biosynthesis. Whereas all previously described members of this general family incorporate a single acyl unit to form the C-25 side chain (the carboxy carbon of this unit becomes C-25), the LL-F28249 macrolides have side chains originating from two acyl units. For (1) and (2) the terminal unit is derived from isobutyrate which is linked *via* the double bond to the propionate-derived unit at C-26. Compounds (3) and (4) have

§ Crystal data for (4): $C_{35}H_{50}O_8$, M = 598.78, orthorhombic, space group $P2_12_12_1$, a = 13.316(4), b = 10.154(2), c = 25.021(5) Å, U = 3383.3 Å³, Z = 4, $D_c = 1.18$ g/cm³, $\lambda(Cu-K_{\alpha}) = 1.54184$ Å, $\mu(Cu-K_{\alpha}) = 6.7$ cm⁻¹, F(000) = 1296. The structure was solved by direct methods and refined by full-matrix least-squares methods. 2209 reflections with $F_o^2 > 3.0\sigma$ (F_o^2) were collected on an Enraf–Nonius CAD4 diffractometer in the range $12 < \theta < 29^\circ$. Refinement led to conventional values of R = 0.069 and $R_w = 0.086$. Atomic co-ordinates, bond lengths and angles, and thermal parameters have been deposited at the Cambridge Crystallographic Data Centre. See Notice to Authors, Issue No. 1. incorporated acetate in place of isobutyrate as the terminal unit. Thus, in general, the polyketide precursors of the LL-F28249 macrolides are one unit longer than those which give rise to other known milbemycin derivatives.

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